



Liqui-PREP™

The Next Generation of Liquid Cytology

Manual Technical Tips

TIP Number: 007

Subject: Liqui-PREP™ Cellular Adhesion

TECHNICAL TIP OVERVIEW:

Cellular adhesion problems for encapsulated Liqui-PREP™ processed cell pellet occur for three reasons:

- Microscope Slides assumed to be clean may have residual cutting oils or detergents which will Interfere with the encapsulated cellular pellet adherence to the glass microscope slide.
- Liqui-PREP™ preparations are not allowed to thoroughly dry before staining.
- Specimen inoculation concentration too heavy.
- “Water wash steps” during staining are applied too vigorously.

NOTE: Technical Tips are intended to be suggested guidelines for handling cytology specimens. The Laboratory Professional may use these guidelines or make modifications as needed to process a given specimen.

Microscope Slides - During the process of making the slides, fine oils are used to cut the glass and detergents are used to clean the glass microscope slides. Generally if one uses high quality glass microscope slides, the oils and detergents are removed prior to packaging. However, even with high quality glass microscope slides, there may be a residual of these oils and detergents on the slides. This situation is easily resolved by washing the glass Microscope slides prior to processing. This procedure is very easy and fast. See **Manual Technical Tip 3 Problematic Glass Microscope Slides**.

In-complete Dried Slides - Complete drying of the Liqui-PREP™ processed slides is an essential step. The drying time for the Liqui-PREP™ processed slides are temperature and humidity dependent and can vary among laboratory. Drying times can be shortened by using heat to rapidly dry the slides as is presented in **Manual Technical Tip 4 Heat Drying of Liqui-PREP™ Processed Slides**.

Specimen Inoculum is too heavy - If the encapsulation inoculum is too heavy, the Liqui-PREP™ Cellular Base Solution will not be able to properly adhere to the slide. Dilute the encapsulation and repeat the slide. This only occurs for users using the in-Centrifuge Tube encapsulation method. This never occurs with the “Working Tube Method”.

H₂O Washing During Staining - With liquid preparation slides, it is not necessary to use the “old fast running water” wash after the Hematoxylin staining step. In replacement of fast running water step, the use of water baths are more than adequate to remove the excess Hematoxylin. (In the LGM laboratory, three water containers - 30 seconds, 30 seconds, and 1 minute, are used prior to continuing onto the 95% Ethanol wash). Alternatively, slides can be washed with a “soft trickle of water” allowed to run into one of the corners of the wash container.

Any questions, contact your local Liqui-PREP™ representative or:

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