



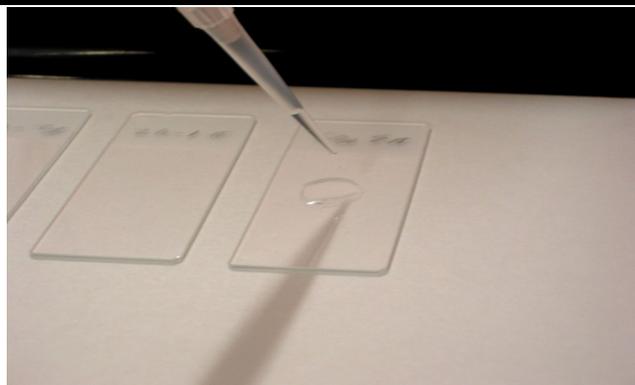
Liqui-PREP[®]

The Next Generation of Liquid Cytology

Technical Tips

Number: 00008

Date: 07/09/22



SUBJECT: Cellular Adhesion/Cell Loss

TECHNICAL TIP OVERVIEW:

Cellular adhesion problems for encapsulated **Liqui-PREP[®]** processed cell pellet occur for four reasons:

1. Microscope Slides, assumed to be clean, may have residual cutting oils or detergents which will interfere with the encapsulated cellular pellet adherence to the glass microscope slide.
2. **Liqui-PREP[®]** preparations are not allowed to thoroughly dry before staining.
3. **Liqui-PREP[®]** preparations are too cellular.
4. “Water wash steps” during staining are applied too vigorously.

TECHNICAL TIP: 00008

1. **Microscope Slides** - During the microscope slide production, fine oils are used to cut the glass and detergents are used to clean the glass microscope slides. Generally if one uses high quality glass microscope slides, the oils and detergents are removed prior to packaging. However, even with high quality glass microscope slides, there may have residuals of these oils and detergents.

This situation is easily resolved by washing the glass Microscope slides prior to processing. This procedure is very easy and fast. See **Technical Tip TT00003 Problem Glass Microscope Slides**.

2. **In-complete Dried Slides** - Complete drying of the **Liqui-PREP[®]** processed slides is an essential step. The drying time for the **Liqui-PREP[®]** processed slides are temperature and humidity dependent, which can vary among laboratory. Drying times can be shortened by using heat to rapidly dry the slides as is presented in **Technical Tip TT00004 Heat Drying of Liqui-PREP[®] Processed Slides**.

3. **Cell Suspension too Cellular** - If the concentration of cells in the **Liqui-PREP[®]** Cellular Base Solution is too cellular, the encapsulation can be overwhelmed causing wash off. Add more **Liqui-PREP[®]** Cellular Base Solution to the “Working Tube”, mix well and make another slide.

4. **H₂O Washing During Staining** - With liquid preparation slides, it is not necessary to use the “old fast running water” wash after the Hematoxylin staining step. In replacement of fast running water step, the use of water baths are more than adequate to remove the excess Hematoxylin. (In the LGM laboratory, three water containers - 30 seconds, 30 seconds, and 1 minute, are used prior to continuing onto the 95% Ethanol wash). Alternatively, slides can be washed with a “soft trickle of water” allowed to run into one of the corners of the wash container.

Any Questions, Contact your local Liqui-PREP[®] Representative :

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